A. **PURPOSE:** The CMI at NCRC allows use of euthanasia room G014 to those investigators whose animals are housed within G012 for the purposes of imaging. To provide for the integrity and well-being of the animals in the imaging facility, a system has been devised that will facilitate the research while minimizing the risk of spreading a contaminant. It is critical that research personnel follow the procedures described in the *Guide for the Care and Use of Laboratory Animals* regarding the transportation of animals in order to reduce any undue stress and risk of zoonoses to and from the animals. The following Standard Operating Procedure (SOP) outlines the steps required to maintain a sanitary imaging environment for the research animals.

B. **PROCEDURES:** Since many of the animals used in the CMI are immunodeficient nude or SCID mice, it is very important that all users with biologically hazardous animals adhere to the strict hygienic guidelines provided in this Standard Operating Procedure (SOP) to prevent cross contamination. The following guidelines have been devised to ensure the likelihood of a zoonotic pathogen being transferred between animals has been minimized. This SOP also details the required procedures for users working with animals harboring chemically hazardous agents that require containment housing. This housing requirement should be detailed in the beginning pages of your approved animal protocol. Please contact Amanda Fair at awelton@umich.edu for further information.
1. Use of euthanasia room G014 is restricted to CMI users and their non-hazardous animals only. If you need to euthanize animals harboring biological or chemical hazards you will need to work within the biological safety cabinet located in containment housing room G013.

2. Specific PPE must be worn when entering or working within the euthanasia room. This includes disposable gown, hair cover and gloves.

3. Before bringing animals into the euthanasia room, transfer the animals into a clean cage using micro-isolation techniques. Please contact ULAM or visit [www.ulam.umich.edu](http://www.ulam.umich.edu) if you need a refresher course or are unsure of the proper micro-isolation techniques.
4. A cart is kept in the hallway to assist in the transfer of cages from the animals rooms to the procedure room. Please note that the transfer cart cannot enter either type of room.

5. Prior to working in the euthanasia room users are responsible for cleaning all counters that will be used. Dry the surfaces with a clean paper towel and dispose of the dirty toweling in a garbage container.

6. When procedures are concluded, liberally spray all work surfaces with Virex disinfectant solution (provided). Leave on for ~10 minutes and wipe clean with a paper towel.

7. Dispose of all soiled paper products in the biological hazard container kept within the room. Tissues/liquids/animals must be disposed of in the appropriate container within the ULAM carcass cooler, they should never be disposed of within a CMI room.

8. All sharp objects such as scalpel blades, hypodermic needles, Pasteur pipettes and, contaminated broken glass etc. must always be disposed of in the approved "sharps bins". USED NEEDLES MUST NOT BE RECAPPED AFTER USE. The needle syringe assembly should be placed promptly in a sharps bin for disposal. Do not leave uncapped needles or sharps on surfaces. Please refer to the OSEH website and the "Guidance on the Use of Sharps" http://www.oseh.umich.edu/sharps_guide.html for further information.